A SHORT, ENANTIOSPECIFIC SYNTHESIS OF THE la-HYDROXYVITAMIN D ENYNE A-RING SYNTHON

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<u>Abstract</u>: The vitamin D A-ring synthon, the enyne 3b, was synthesized in 5 steps (37% overall yield) from (S)-(+)-carvone (5). The key step was the SmI_2 -Pd⁰ mediated transformation of epoxypropargyl ester 6 to 3b.

It has been recently established that $1\alpha,25-dihydroxyvitamin D_3 [1\alpha,25-(OH)_2D_3]$ (1), the hormonal form of vitamin D₃, is able to suppress proliferation and induce differentiation in human myeloid leukemia cells.¹ More classically, this hormone is considered to be the most potent stimulator of calcitropic effects known.² These findings have stimulated efforts towards the search for analogues of $1\alpha,25-(OH)_2D_3$ that, while retaining potent cell differentiating and anti-proliferative activity, do not cause the toxic hypercalcemia associated with the normal calcitropic effects of $1\alpha,25-(OH)_2D_3$.



Lythgoe's convergent approach to the synthesis of vitamin D derivatives makes use of the coupling of an A-ring enyne with an appropriate CD-ring fragment.³ Ongoing investigations in our laboratories have led to the utilization of a [1,5]-sigmatropic H-shift of vinylallenes of the type 2 to generate the triene portion of vitamin D.⁴ The hydroxyenyne 3 should be a suitable A-ring fragment for the generation of such vinylallene systems⁵ derived from the

coupling of a suitably protected form of ${\bf 3}$ and a CD-ring fragment 4.

This A-ring synthon 3 was first prepared in 12 steps and 3% overall yield by Lythgoe who used it in a synthesis of 1α ,25-(OH)₂D₃.⁶ More recently, Castedo et al have reported⁷a a more efficient synthesis (11 steps, 10% overall yield) of 3 and the synthesis of a closely related substance has been independently reported by the Hoffmann-La Roche group⁷b and by Desmaele and Tanier⁷c.

We now report an expeditious, enantiospecific synthesis of 3 starting from S-(+)-carvone 5, a starting material also utilized by Castedo et al^{7a} and the Roche group.^{7b} The key step in the synthesis is a novel SmI₂-promoted reductive elimination of the epoxypropargyl acetate 6 with concomitant ring opening of the epoxide moiety to afford the enynol 3b.



Our synthesis makes use of the known stereoselective epoxidation of S-(+)-carvone to give cis-carvone epoxide 7 (89%).⁸ Addition of lithium acetylide⁹ (THF, -78 °C) to 7 proceeded highly stereoselectively to afford the propargyl alcohol 8^{10} (81%) (tentatively assigned the stereochemistry shown^{11,12}).



After acetylation of the hydroxyl group (acetic anhydride, Et₃N, DMAP, 25 °C, 87%) to give $9,^{13}$ ozonolysis (0₃, CH₂Cl₂, MeOH, -78 °C) of the isopropenyl side chain followed by direct acylation of 10 (p-nitrobenzoyl chloride, CH₂Cl₂, pyridine, 0 °C) and in situ Criegee rearrangement¹⁴ (40 °C) of the resulting methoxyperoxyester 11 afforded the key diacetate $6,^{15}$ in 70% overall yield from the monoacetate 9.

The crucial reductive elimination of 6 to 3b was effected using the method recently developed by Inanaga for the conversion of propargyl acetates into allenes.^{5b} Under slightly modified conditions [2 eq SmI₂, Pd(PPh₃)₄, THF, 25 °C, absence of a proton source¹⁶) the presumed palladium complex intermediate 12 underwent not only reduction but also epoxide ring opening to cleanly afford the hydroxyenyne $3b^{17}$ in 83% yield [five steps, 37% overall yield from (S)-(+)-carvone].

The use of the A-ring synthon 3 in the synthesis of $l\alpha$,25-(OH)₂D₃ analogues suitable for biological assay as well as the development of further uses in this context of the new reductive elimination-epoxide ring opening process are presently in progress.

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References

- (1) (a) Mangelsdorf, D.J.; Koeffler, H.P.; Donaldson, C.A.; Pike, J.W.; Haussler, M.R. J. Cell. Biol., 1984, 98, 391. (b) Honma, Y.; Hozumi, M.; Abe, E.; Konno, K.; Fukushima, M.; Hata, S.; Nishii, Y.; DeLuca, H.F.; Suda, T. Proc. Natl. Acad. Sci. USA 1983, 80, 201. (c) Koeffler, H.P.; Amatruda, T.; Ikekawa, N.; Kobayashi, Y.; DeLuca, H.F. Cancer Res. 1984, 44, 5624. (d) MacLaughlin, J.A.; Gange, W.; Taylor, D.; Smith, E.; Holick, M.F. Proc. Natl. Acad. Sci. USA 1985, 82, 5409.
- (2) (a) Norman, A.W., "Vitamin D, the Calcium Homeostatic Steroid Hormone", Academic Press, New York, 1979. (b) DeLuca, H.F.; Paaren, H.E.; Schnoes, H.K. Topics in Current Chemistry, 1979, 83, 1. (c) Norman, A.W.; Roth, J.; Orci, L. Endocrine Reviews, 1982, 3, 331. (d) Henry, H.L.; Norman, A.W. Ann. Rev. Nutrition, 1984, 4, 493.
- (3) (a) Lythgoe, B. <u>Chem. Soc. Rev.</u>, 1981, 449. (b) Pardo, R.; Santelli, M. <u>Bull. Soc.</u> <u>Chim. Fr.</u>, 1985, 98. (c) Sardina, F.J.; Mouriño, A.; Castedo, L. <u>J. Org. Chem.</u>, 1986, <u>51</u>, 1264. (d) Mascareñas, J.L.; Mouriño, A.; Castedo, L. <u>J. Org. Chem.</u>, 1986, <u>51</u>, 1269. (e) Baggiolini, E.G.; Iacobelli, J.A.; Hennessy, B.M.; Batchco, A.D.; Sereno, J.F.; Uskokovic, M.R. <u>J. Org. Chem.</u>, 1986, <u>51</u>, 3098.
- (4) Okamura, W.H. Acc. Chem. Res., 1983, 16, 81.
- (5) See: (a) Schuster, H.F.; Coppola, G.M., "Allenes in Organic Synthesis", Wiley & Sons, New York, 1984, pp. 12, 89. (b) Tabuchi, T.; Inanaga, J.; Yamaguchi, M. <u>Tetrahedron</u> <u>Lett.</u>, 1986, <u>27</u>, 5237.
- (6) Harrison, R.G.; Lythgoe, B.; Wright, P.W. J. Chem. Soc. Perkin Trans. 1, 1974, 2654.
- (7) (a) Castedo, L.; Mascareñas, J.L.; Mouriño, A. <u>Tetrahedron Lett.</u>, 1987, <u>28</u>, 2099. (b) Baggiolini, E.G.; Hennessy, B.M., Iacobelli, J.A.; Uskokovic, M.R. <u>Tetrahedron Lett.</u>, 1987, <u>28</u>, 2095. (c) Desmaele, D; Tanier, S. <u>Tetrahedron Lett.</u>, 1985, <u>26</u>, 4941.
- (8) Klein, E.; Ohloff, G. <u>Tetrahedron</u>, 1963, <u>19</u>, 1091.

- (9) Midland, M.M. J. Org. Chem., 1975, 40, 2250.
- (10) Data for 8: mp 55-56°C; $[\alpha]_D^{25}$ 0° (c 2.0, CHC23); ¹H NMR (200 MHz, CDC23) δ 1.5-1.8 (2H, m), 1.60 (3H, s), 1.71 (3H, s), 2.0-2.3 (3H, m), 2.58 (1H, s), 2.97 (1H, s), 3.36 (1H, m), 4.72 (1H, br s), 4.77 (1H, m). All new compounds exhibited satisfactory spectral and mass spectral analytical data. Additional selected spectroscopic data are presented in footnotes 13, 15 and 17.
- (11) The assignment was made on the basis of preferential attack at the less hindered face of the epoxyketone, i.e. opposite to the 10β -methyl¹² group. For a discussion on the stereochemistry of hydride attack on α -ketoepoxides, see Chautemps, P.; Pierre, J.L. Tetrahedron, 1976, 32, 549.
- (12) The steroidal numbering system is used wherein the methyl group corresponds to C_{10} and is attached to C_{10} .
- (13) Data for 9: bp (Kugelrohr) 110° C/1.4 mm Hg; $[\alpha]_D^{25} + 0.8^{\circ}$ (c 1.8, CHCl₃); ¹H NMR (200 MHz, CDCl₃) δ 1.5-1.7 (2H, m), 1.66 (3H, s), 1.67 (3H, s), 2.03 (3H, s), 2.1-2.3 (3H, m), 3.0 (1H, m), 4.66 (1H, br s), 4.70 (1H, br s).
- (14) Schreiber, S.L.; Liew, W.F. Tetrahedron Lett., 1983, 24, 2363.
- (15) Data for 6: mp 115-116°C; $[\alpha]_D^{25} 19.0°$ (c 0.5, CHCl₃); ¹H NMR (200 MHz, CDCl₃) δ 1.61 (3H, s), 1.94 (1H, dd, J = 14.2, 3.4 Hz), 2.03 (3H, s), 2.10 (3H, s), 1.9-2.1 (1H, m), 2.35 (1H, dd, J = 15.8, 6.1 Hz), 2.70 (1H, s), 2.83 (1H, ddd, J = 14.2, 5.4, 1.0 Hz), 3.15 (1H, d, J = 3.4 Hz), 4.97 (1H, m).
- (16) Studies on the model system 13 showed that in addition to the desired enyne 14, unidentified side-products were obtained if, following Inanaga's conditions, a proton source was used.



(17) Data for **3b**: oil; $[\alpha]_D^{25} - 101.3^\circ$ (c 3.0, CHCl₃); ¹H NMR (200 MHz, CDCl₃) $\delta 1.8-2.0$ (8H, m), 2.1 (1H, m), 2.5 (1H, m), 3.06 (1H, s), 3.3 (1H, br), 4.1 (1H, br), 5.05 (1H, m). Further characterized as the bis(trimethylsilyl)ether **3c**, previously reported by Lythgoe.⁶ Data for **3c**: oil, $[\alpha]_D^{25} -102.5^\circ$ (c 0.4, CHCl₃) [lit.⁶ $[\alpha]_D^{22}$ (CHCl₃) - 90°]; ¹H NMR⁶ (200 MHz, CDCl₃) δ 0.13 (9H, s), 0.17 (9H, s), 1.6-1.9 (2H, m), 1.92 (3H, br s), 2.1 (1H, m), 2.4 (1H, dd, J = 16.6, 4.9 Hz), 3.05 (1H, s), 4.0-4.2 (2H, m).

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